

# THE EUKARYOTE CELL INTERACTION WITH DOPED TiO<sub>2</sub> NANOPARTICLES

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**Abstract.** *The experiment was performed in Mus musculus, being analyzed the liver ultra-structural features, as well as the nanoparticles interaction (single TiO<sub>2</sub> nanoparticles of 10-20 nm, or conjugate with a metal, TiO<sub>2</sub>-Fe, TiO<sub>2</sub>-Pt or TiO<sub>2</sub>-Ag) with the hepatic cell. The small nanoparticles can penetrate in hepatocyte, their effect being dependent on the conjugate metal kinds. The TiO<sub>2</sub>-Pt and TiO<sub>2</sub>-Ag nanoparticles induced reversible effect, while the TiO<sub>2</sub>-Fe nanoparticles alter the hepatocyte ultra-structure. Depending on the metal kinds, the TiO<sub>2</sub>-Me nanoparticles can be in relation with smooth and rough endoplasmic reticulum and with mitochondria, or in lipid droplets, being also present in the Kupffer cell.*

## 1. Introduction

Initially TiO<sub>2</sub> nanoparticles were considered an inert material, because it is formed of very small size. Subsequent, was point that the TiO<sub>2</sub> nanoparticles can induce lesions at the chromosomes and DNA, as well as at the ultrastructural level. They can migrate in different organs, and induce oxidative stress and cell death. As result of the studies performed by International Agency for Research on Cancer (IARC), was re-revaluated the TiO<sub>2</sub> toxicity, they being moved in the **2B** group of materials, as possible carcinogen for men [1]. The biological reactivity of the TiO<sub>2</sub> nanoparticles is dependent on different features: their crystallization shape (anatase, rutile or brookite), amount, size, a/o. Recently, were obtained TiO<sub>2</sub> nanoparticles doped with different chemical elements, as well as nanoparticles included in liposome. From the noble metals, the mostly used were silver, gold and platinum, as well as other metals (iron, copper, a/o). The noble metals are resistant at oxidation process. In many studies, the silver was deposited on TiO<sub>2</sub> surface, being more reactive than gold or platinum, having the photocatalytic effect proper to TiO<sub>2</sub> nanoparticles, as well as the ability to prevent bacteria development [2].

In the human or animal body, the research was performed to establish an eventual cytotoxic effect depending on different parameters: the nanoparticle size [3], their single presence or doped with other element [4], activation or not by UV rays [5], [6], encapsulated or not in different bioactive substances, a/o. The cyto- and genotoxicity analysis of the ultrafine TiO<sub>2</sub> nanoparticles, in human lymphoblast cell, evidenced that these can induce cyto- and genotoxicity in a significant mode [7]. The analysis of the biological effects of three kinds of nano-TiO<sub>2</sub> of different size (10-20 nm anatase; 50-60 nm anatase and 50-60 nm rutile) on DNA and cell ultrastructure of the 293T and CHO cells, reveal that the all three kinds of nano-TiO<sub>2</sub> manifest a higher toxic effect on tumor cells than on normal cells [8]. Also, the nano-TiO<sub>2</sub> from anatase kinds, were observed in the cytoplasm of CHO cells, indifferent on this size (50-60 nm or 10-20 nm).

The biological investigation point out different biological effects of the TiO<sub>2</sub>-Me nanoparticles, depending on the metal type (silver, golden, copper or platinum). Thus, the use of the TiO<sub>2</sub>-Pt and TiO<sub>2</sub>-Au nanoparticles, are preferable in comparison with TiO<sub>2</sub>-Ag or TiO<sub>2</sub>-Cu doped nanoparticles [9].

There are a few studies regarding the TiO<sub>2</sub> nanoparticles interaction with the eukaryote cell. In a synthesis from 2006 [10], there wasn't present any paper regarding the interaction between the eukaryote cell with the TiO<sub>2</sub> nanoparticles. In previously researches, our groups point out the relation between TiO<sub>2</sub> nanoparticles with eukaryote animal cell [11], or vegetal cell [12]. In other paper [8], was underlined that the nano-TiO<sub>2</sub> from anatase kinds, were observed in the cytoplasm of CHO cells, indifferent on this size (50-60 nm or 10-20 nm).

In this paper the interaction of titanium dioxide nanoparticles with animal cell (hepatocyte from *Mus musculus*), depending on the chelate metal (single or chelate with silver, platinum, or iron) was analyzed.

## **2. Materials and methods**

### **2.1. The preparation of titanium dioxide nanoparticles**

Undoped and doped titanium dioxide nanocrystals were synthesized by the sol-gel route, using the precursors: titanium isopropoxide, isopropyl alcohol, distilled water, nitric acid, hexachloroplatinate acid, silver nitrate and ferrous nitrate. Over 30 ml of isopropyl alcohol, 5 ml of titanium isopropoxide were added in drops, under continuous stirring with the magnetic stirrer. After a few minutes of stirring, distilled water was added, continuously controlling the solution pH with nitric acid in order to avoid the precipitation. In the case of platinum-doped and iron-doped TiO<sub>2</sub> ions, after the adjustment of the pH (pH=2.5 for Pt and Ag, pH=5 for Fe), the previously prepared solutions, of were added hexachloroplatinate acid for doped with Pt, silver nitrate for doped with Ag and ferrous nitrate for doped with Fe under continuous stirring. In all cases, the gel was dried and washed in order to remove the secondary reaction products. The calcinations was achieved in the oven, at a temperature of 250°C for undoped TiO<sub>2</sub> and 500°C for Pt, Ag or Fe-doped TiO<sub>2</sub>, for 3 hours.

The obtained materials were characterized by X-ray diffraction (XRD), scanning electron microscopy (SEM) and energy dispersive X-ray analysis (EDAX). XRD spectra were recorded at room temperature with a BRUKER D8 ADVANCE X-ray diffractometer using Cu K $\alpha$  radiation in  $\theta$ :2 $\theta$  configuration. The SEM images were made in an Inspect S scanning electron microscope coupled with EDAX device.

### **2.2. Biological material**

The experiments were performed on young females of *Mus musculus*, intra-peritoneal injected with a suspension of titanium dioxide. The animals were intraperitoneal injected (five injections of 0.5 ml each, one two days) with a 0.01% TiO<sub>2</sub> or TiO<sub>2</sub>-Me suspension. A day after the last injection, the animals were sacrificed through the carotid artery section, and their liver were used for the electron microscopy investigations.

### **2.3. Electron microscopy investigation**

The pieces of about 1 mm<sup>3</sup> of liver, were prefixed in a 2.7% glutaraldehyde solution (2 ½ hours), postfixed in a 1% Millonig solution (1 ½ hour) and then included in Epon 812. The seriated sections of about 60 nm thickness, were contrasted with uranyl acetate and lead citrate, and then analyzed at a TEM JEM JEOL-1010 electron microscope in Electron Microscopy Centre, *Babes-Bolyai* University (Cluj-Napoca, Romania).

### 3. Results and discussions

#### 3.1. TiO<sub>2</sub> particles feature

The XRD patterns (Fig. 1) analyses present the crystallization as *anatase* form of the undoped / Pt, Ag or Fe doped TiO<sub>2</sub>, even if the calcinations temperatures for the TiO<sub>2</sub> doping surpass the value of 500°C. The presence of the dopant in the crystalline network of the titanium dioxide tripped the phase transition from *anatase* to *rutile*. From the diffraction spectra it is noticed that the dopants did not present separate peaks, which means that this is distributed uniformly in the crystalline network.

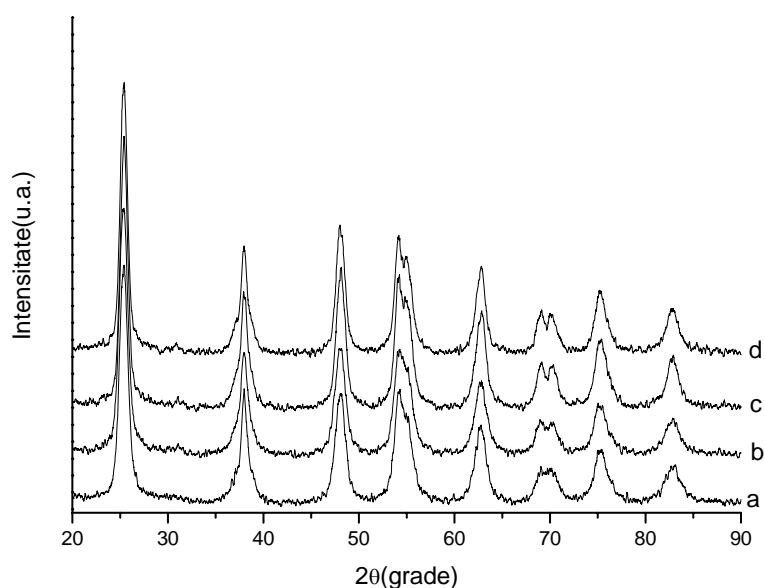


Fig. 1. XRD pattern for: (a) TiO<sub>2</sub> undoped; (b) TiO<sub>2</sub>-Fe; (c) TiO<sub>2</sub>-Pt; (d) TiO<sub>2</sub>-Ag.

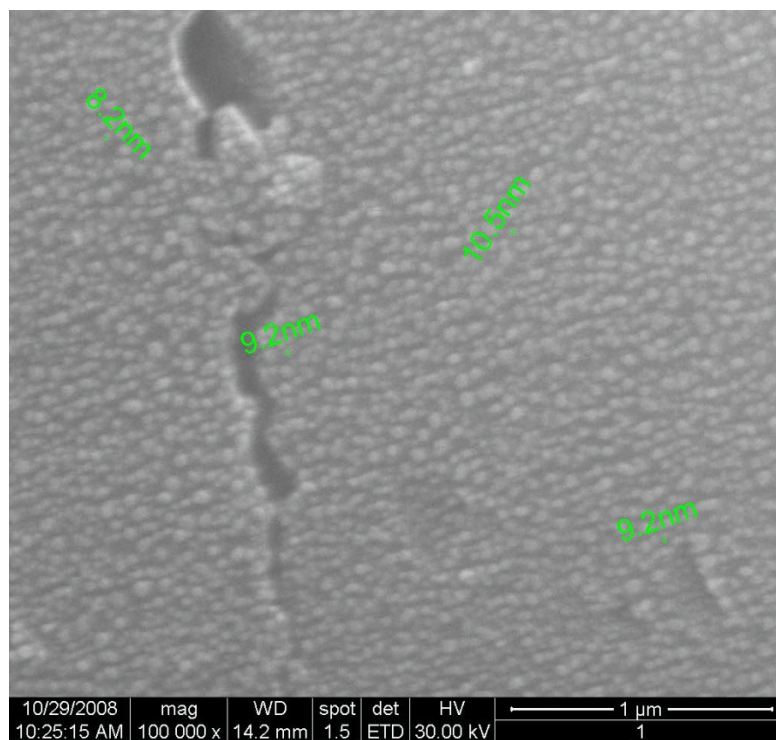


Fig. 2. SEM morphology for  $\text{TiO}_2$  undoped.

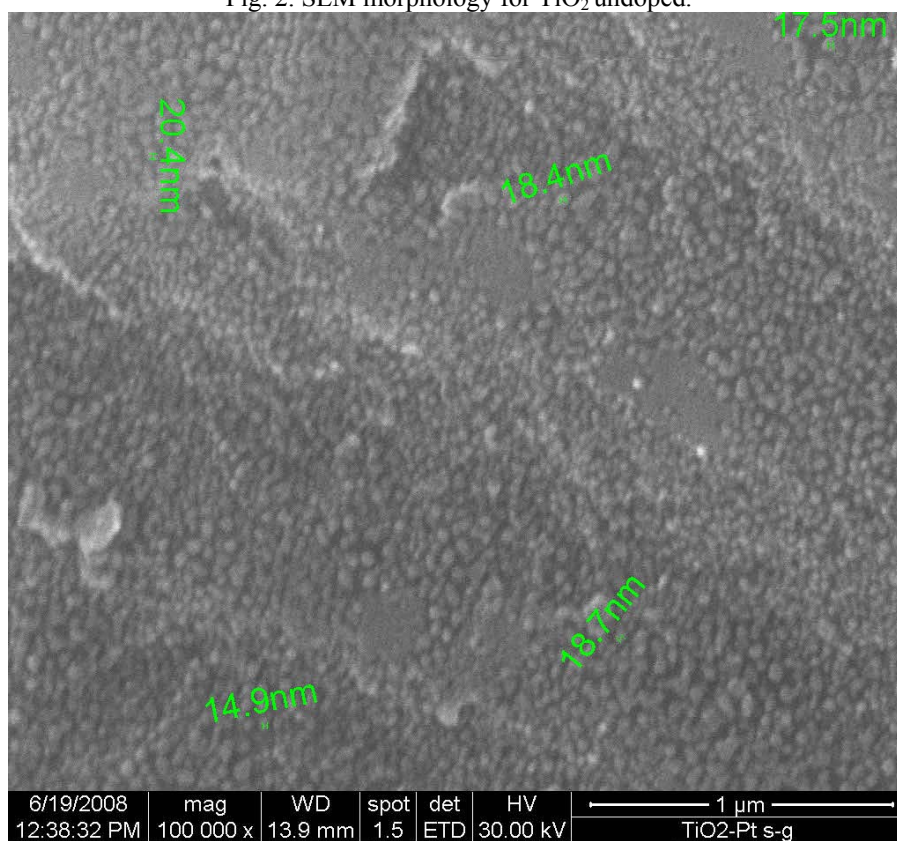


Fig. 3. SEM morphology for  $\text{TiO}_2$ -Pt

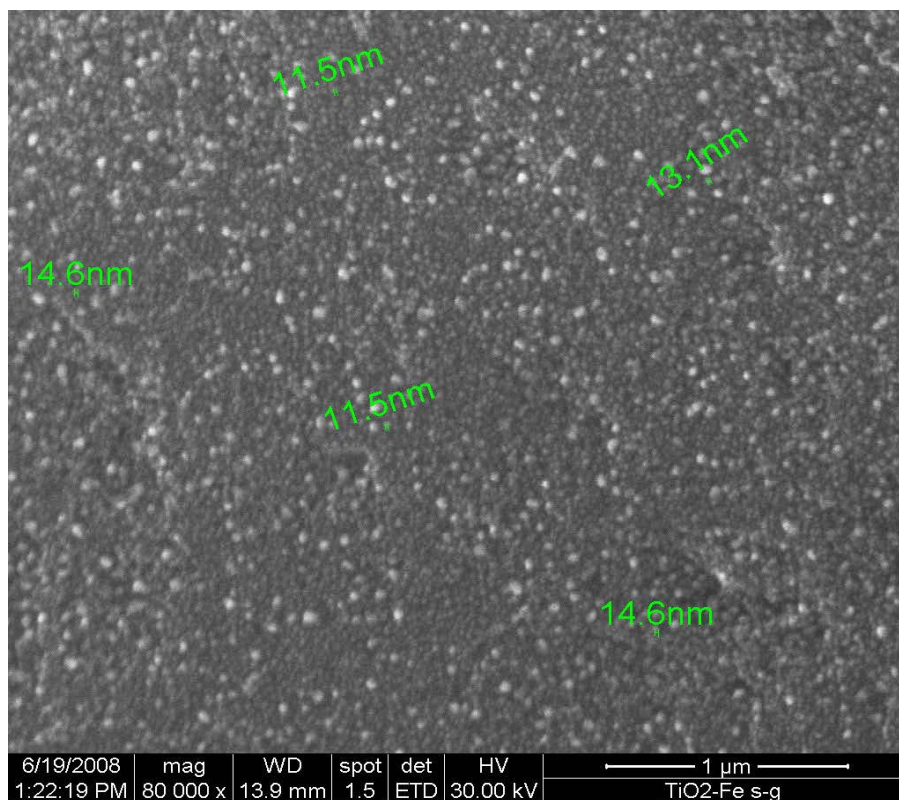


Fig. 4. SEM morphology for TiO<sub>2</sub>-Fe

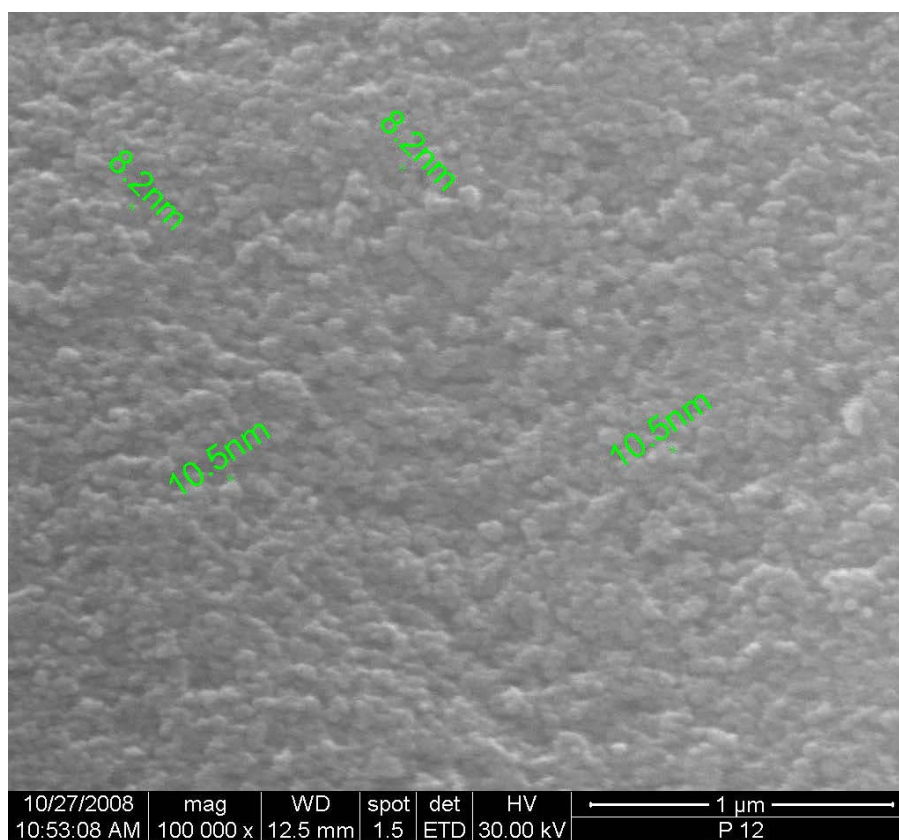


Fig. 5. SEM morphology for TiO<sub>2</sub>-Ag

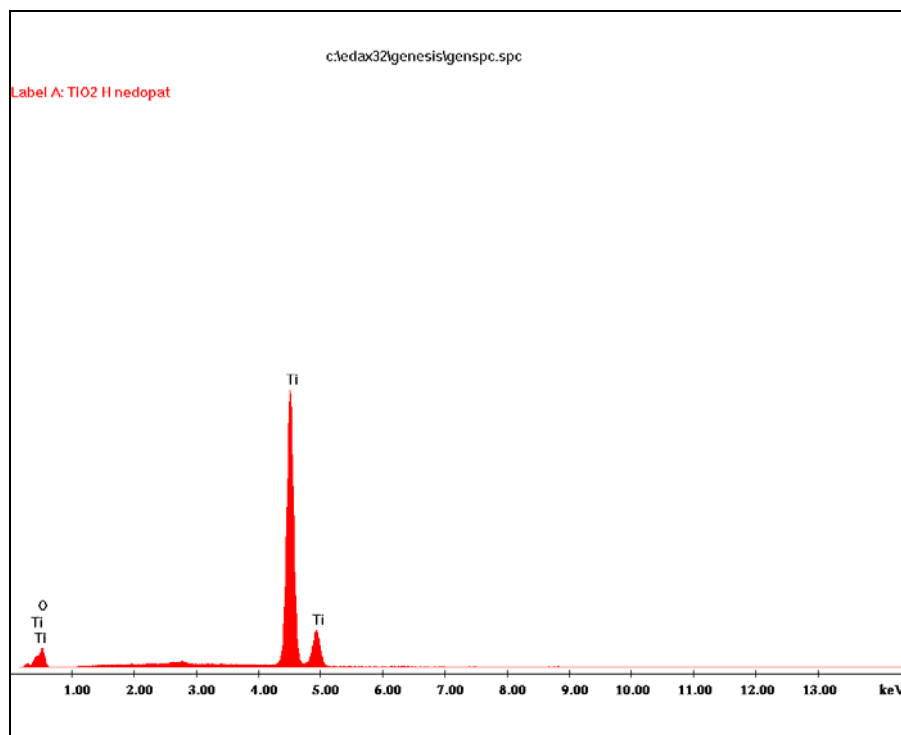


Fig. 6. EDX spectrum for elemental analysis of undoped  $\text{TiO}_2$

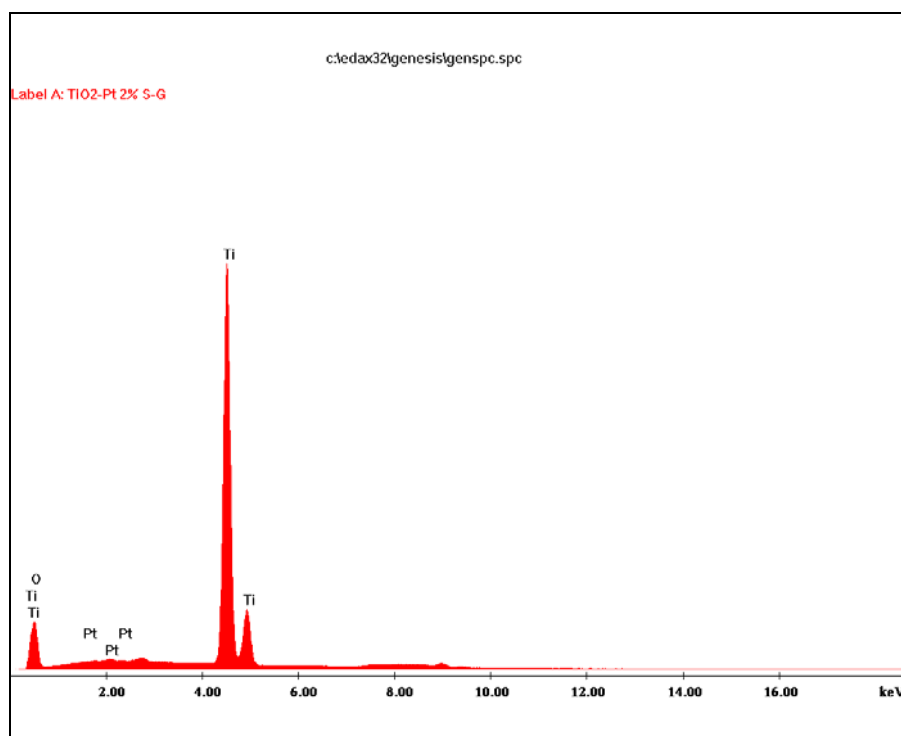


Fig. 7. EDX analysis for elemental analysis of  $\text{TiO}_2$ -Pt



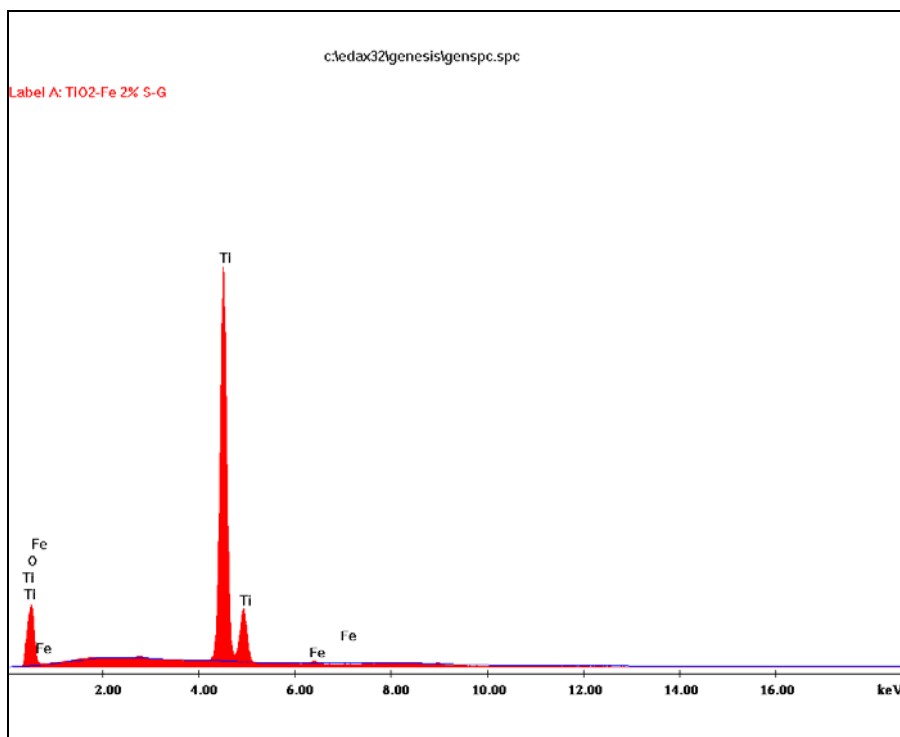


Fig. 8. EDX analysis for elemental analysis of TiO<sub>2</sub>-Fe

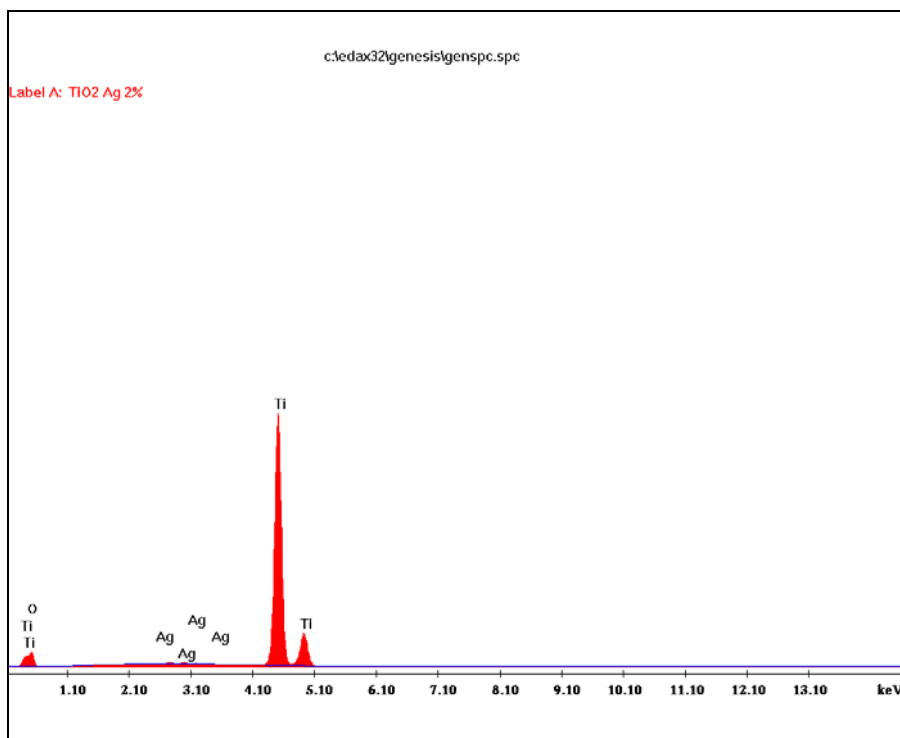


Fig. 9. EDX analysis for elemental analysis of TiO<sub>2</sub>-Ag

From the surface morphology (SEM) it can be observed that the TiO<sub>2</sub>, as well as TiO<sub>2</sub>-Pt, TiO<sub>2</sub>-Ag and TiO<sub>2</sub>-Fe nanospheres dimensions range between 10-20 nm, having a spherical shape, indifferent of the chelated metal (Figs. 2 - 5). EDAX analysis presents undoped TiO<sub>2</sub> nanoparticles, as well as the nanoparticles with Pt, Ag and Fe ions in titanium dioxide structure (Figs. 6 - 9).

### 3.2. TiO<sub>2</sub> nanoparticles interaction with the eukaryote cell

**Normal liver ultrastructure.** The liver has a normal ultrastructure, similar to other reported data [13], [14]. The hepatocytes present one spherical nucleus (sometimes two), with a regulated outline and heterochromatine disposed in a small electrondense masses, usually under inner envelope (Fig. 10). In cytoplasm there are numerous mitochondria spherical or slightly elongated with cristae disposed transversally, rough endoplasmic reticulum well represented, numerous glycogen particles (Fig. 11), Golgi complex, smooth endoplasmic reticulum (SER) with a discrete presence, having a bigger concentration in the areas with glycogen, a/o. The lipid droplets are very rarely in cytoplasm, as small anelectrondense granules, especially at the vascular pole of the hepatocyte in the vicinity of Ito cell (Fig. 12).

The Kupffer cells are in a normal activity (Fig. 13), having accumulated a small amount of phagocytated material.

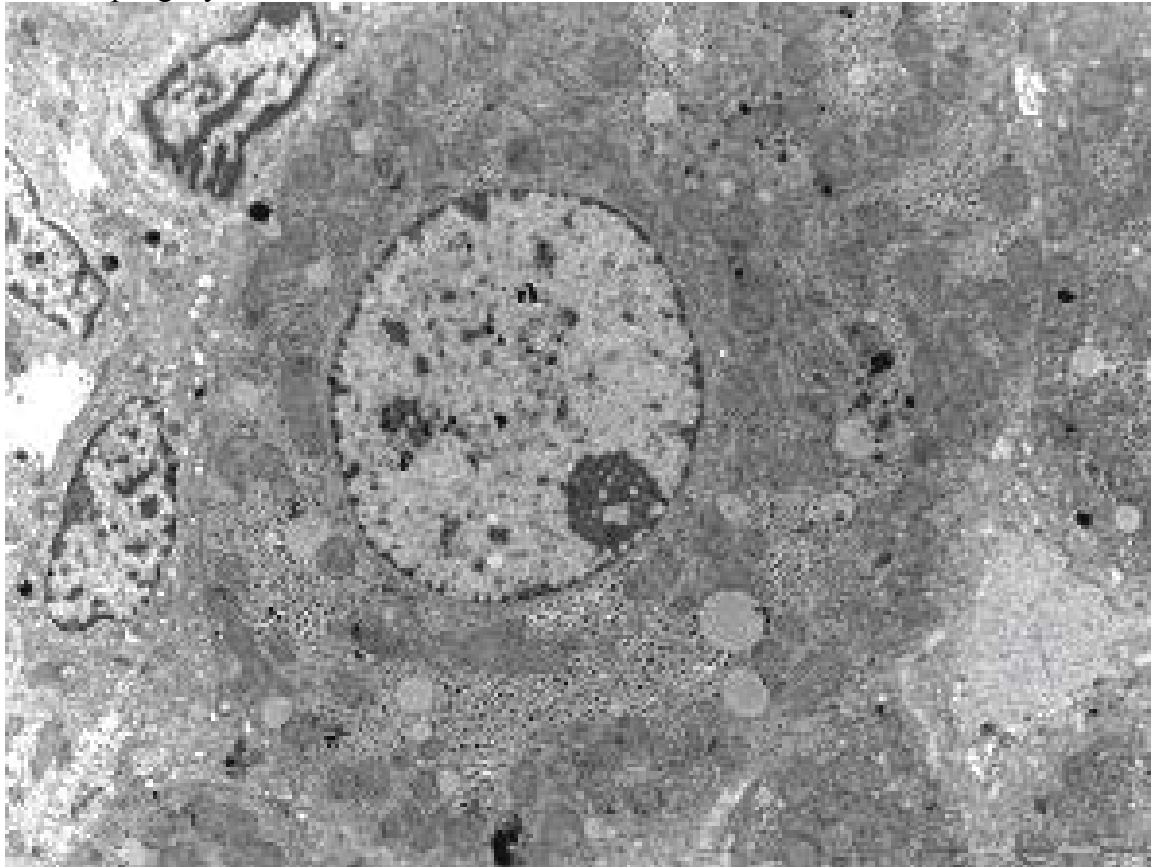


Fig. 10. Control. Hepatocyte with the normal ultrastructure.



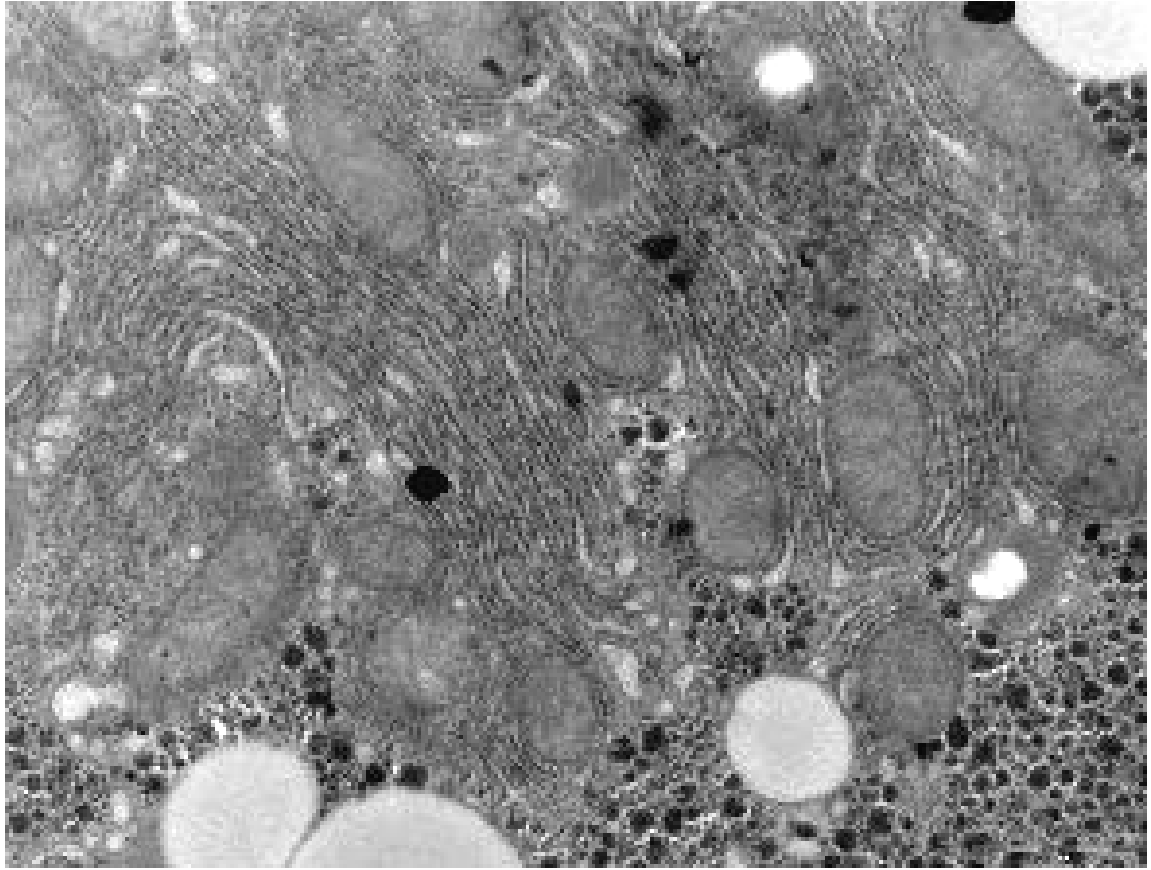


Fig. 11. Control. Cytoplasm with cellular organelles and glycogen

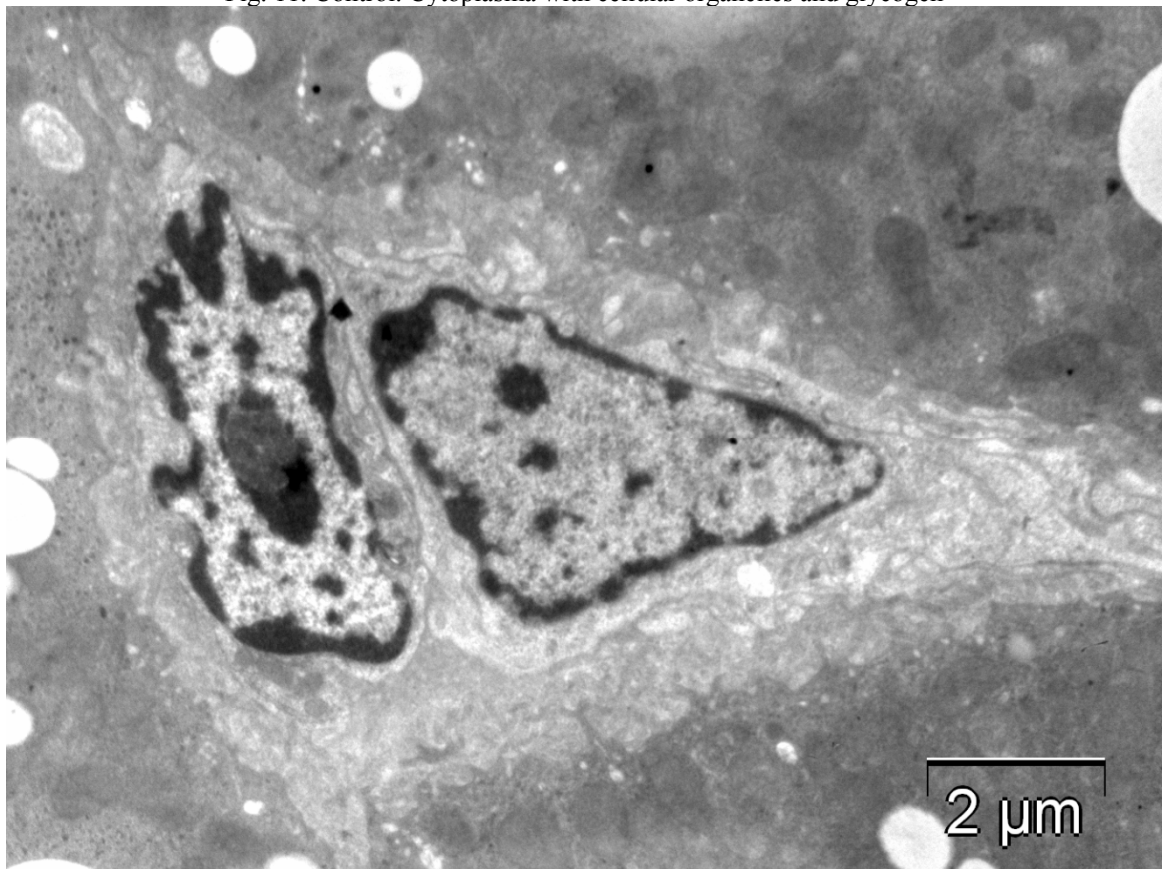


Fig. 12. Control. Vascular pole of hepatocytes and Ito cell.

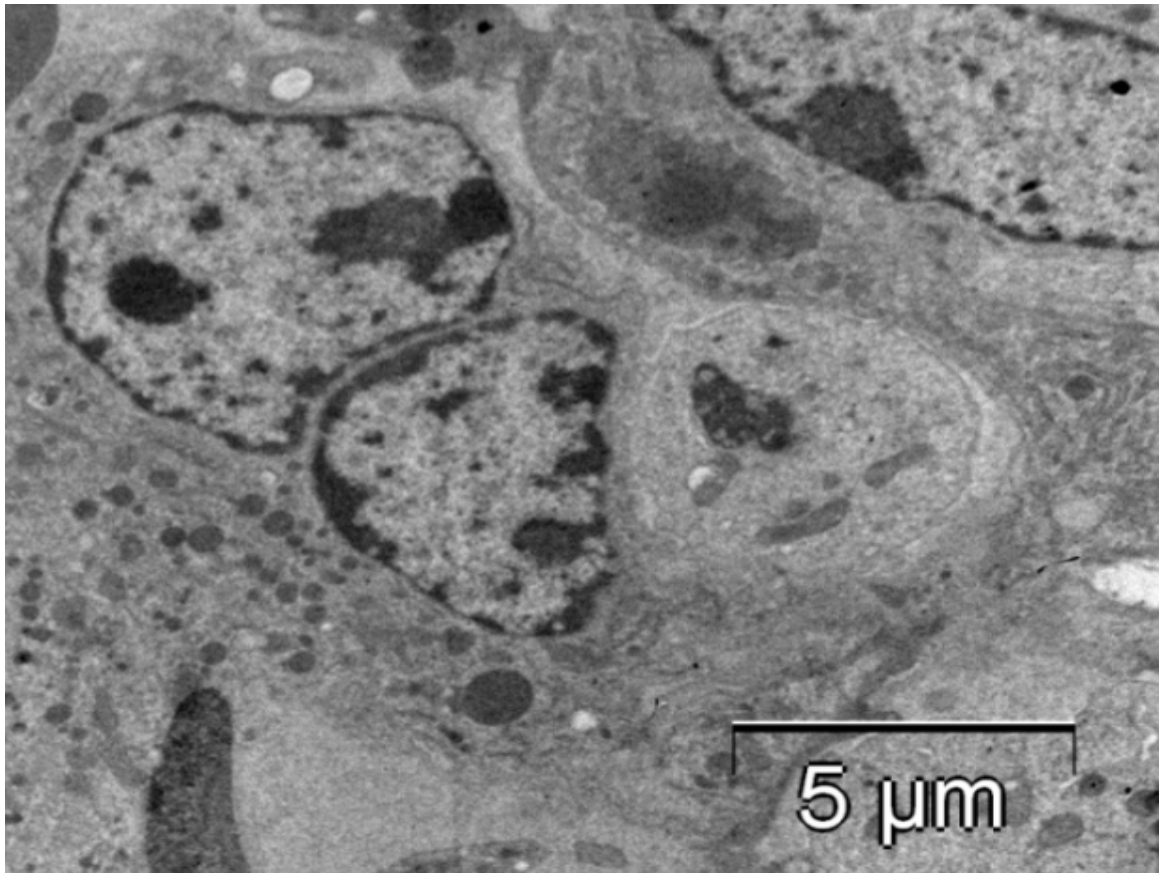


Fig. 13. Control. Kupffer cell in activity.

#### **Undoped TiO<sub>2</sub> nanoparticles effect.**

The presence of undoped TiO<sub>2</sub> nanoparticles, has a drastic affect under the metabolism and ultrastructure of the hepatocyte (Figs. 14-17). The nucleus, usually has 3-4 nucleoli, and an unregulated outline with invaginations, the heterochromatin having a parietal disposition, or spread as fine blocks in euchromatine (Fig. 14). The mitochondria are swelled, with matrix and crista rarefied, sometimes being in amitotic division (Fig. 15). The TiO<sub>2</sub> nanoparticles are accumulated in a big amount in the lipid droplets which became electrondense (fig. 15). Their transit from hepatocyte is practically absent, because they are absent in the Ito cell (Fig. 17). The glycogen microparticles are absent in the cytoplasm of hepatocytes. The cell response at the stress factor, is represented through presence, in a great number, of the vesicles of the smooth endoplasmic reticulum (SER), for counteract of the TiO<sub>2</sub> negative effect (Fig. 16). Also, the Kupffer cell is in metabolic activity.

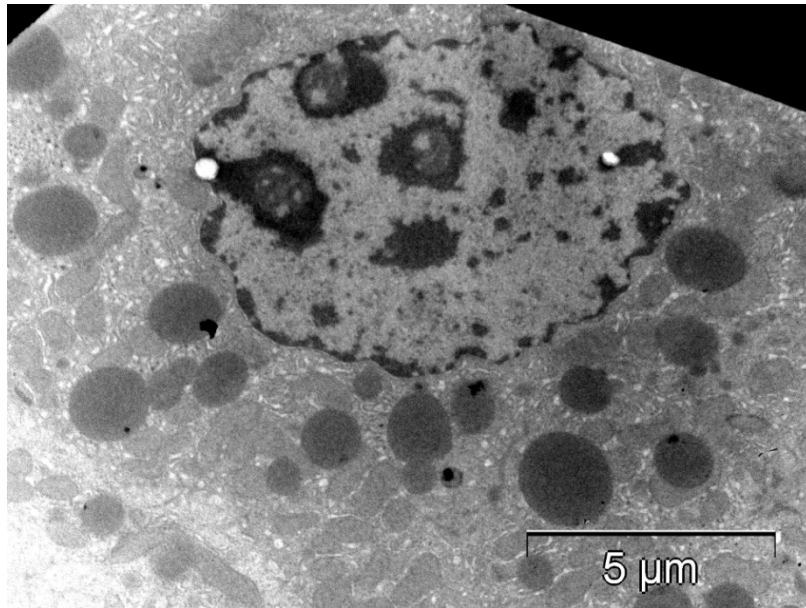


Fig. 14.  $\text{TiO}_2$ . Atered nucleus and many SER vesicles.

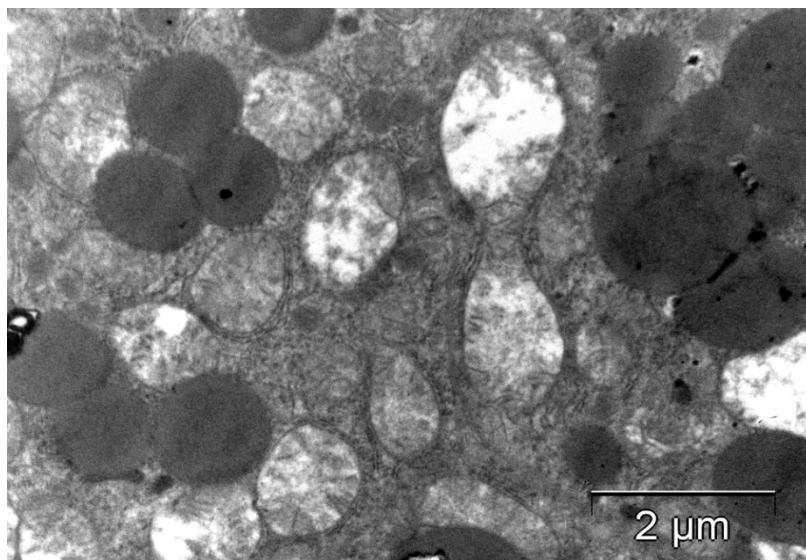


Fig. 15.  $\text{TiO}_2$ . Affected mitochondria, in amitotic division

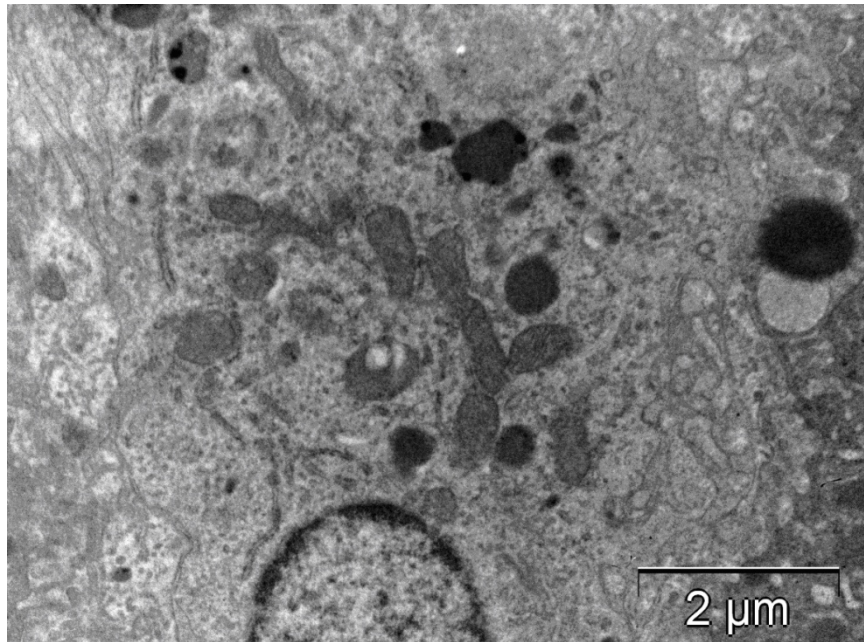


Fig. 16.  $\text{TiO}_2$ . The cell reacts to stress factor.

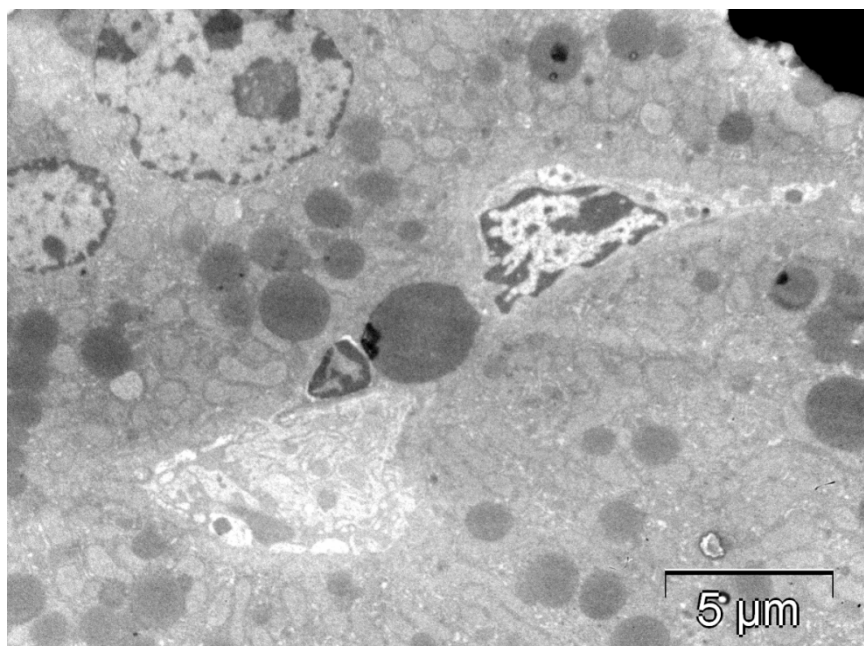


Fig. 17.  $\text{TiO}_2$ . Ito cell, without lipid droplets.

### **The $\text{TiO}_2$ – Ag nanoparticles effect.**

The  $\text{TiO}_2$ -Ag nanoparticles induce reversible modifications in hepatocytes. The nucleus, with 1-3 nucleoli, presents the heterochromatin with parietal disposition or as fine blocks in its inner (Fig. 18). As result of the exogenous nanoparticles presence, the lipid metabolism is altered, hepatocytes having a big amount of lipid droplets (Fig. 18). Lipid droplets contain a moderate amount of  $\text{TiO}_2$ -Ag nanoparticles (Fig. 18). In some mitochondria, the  $\text{TiO}_2$ -Ag nanoparticles are massive accumulated (Fig. 19). As response of this stress factor, the vesicles of smooth endoplasmic reticulum is well represented in proximity (Fig. 19) Also is present Golgi complex. Practically, the glycogen microparticles are absent from hepatocytes.

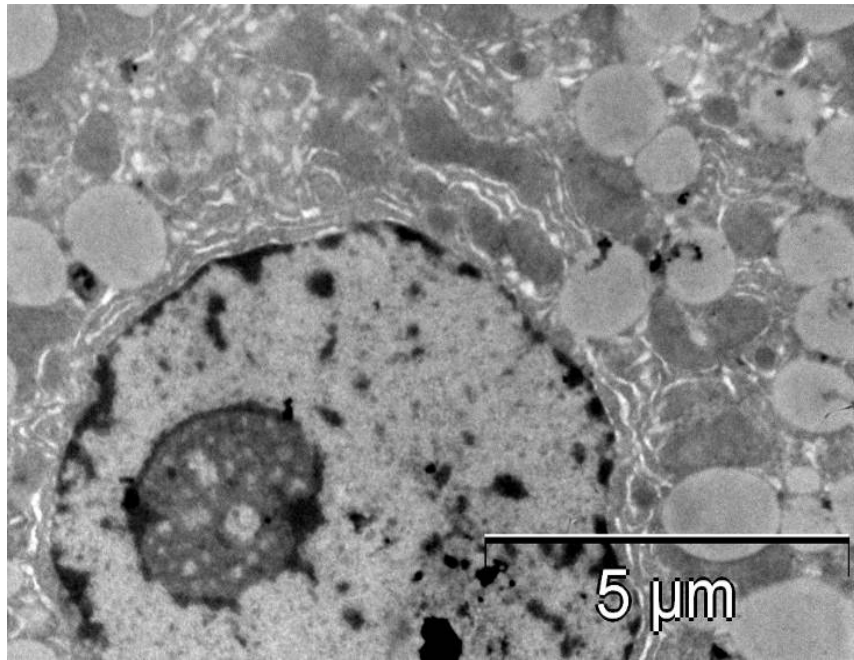


Fig. 18. TiO<sub>2</sub>-Ag. Reversible alterations of the hepatocyte ultrastructure.

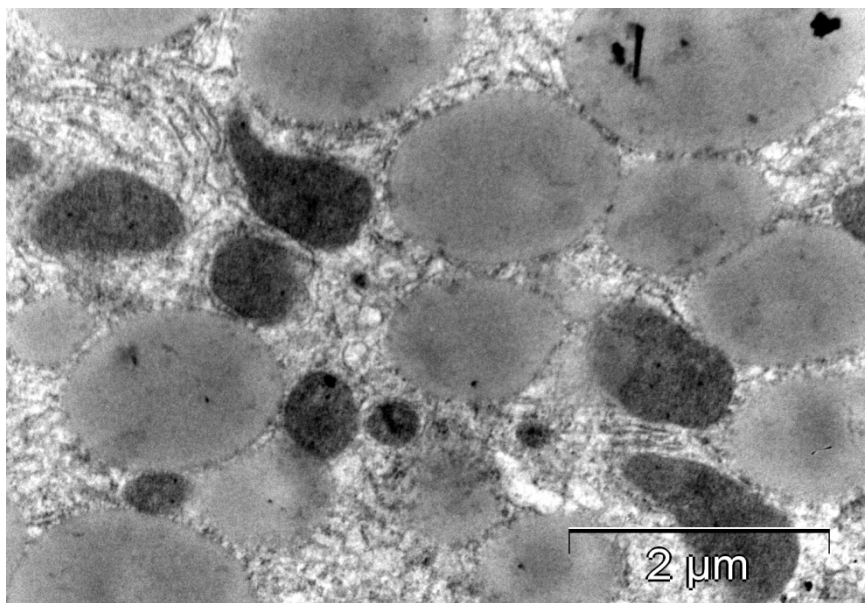


Fig. 19. TiO<sub>2</sub>-Ag. Lipid droplets and mitochondria with TiO<sub>2</sub>-Ag nanoparticles incorporated

Other reported researches [13] was evidenced an enhanced photocatalytic reactivity, in comparison with undoped TiO<sub>2</sub> nanoparticles, and the doped TiO<sub>2</sub> nanoparticles not induced severe alterations in hepatocyte, similar to those induced by undoped TiO<sub>2</sub> nanoparticles.

#### **The TiO<sub>2</sub> – Pt nanoparticles effect.**

The TiO<sub>2</sub>-Pt nanoparticles induced minor and reversible ultrastructural modifications. Some nuclei has an undulated outline (Fig. 20), the profiles of rough endoplasmic reticulum (RER) are slightly dilated and the vesicles of the smooth endoplasmic reticulum are slightly proliferated (Fig. 21).

Also, the hepatocytes do not present collagen fibers and cellular destructions. The mitochondria present a normal ultrastructure (Fig. 21). The small TiO<sub>2</sub>-Pt nanoparticles



penetrate in hepatocytes under shape of electrondense masses, being in strong relation with rough endoplasmic reticulum profiles, smooth endoplasmic reticulum vesicles and especially with mitochondria (Figs. 22, 23). There are mitochondria with a different amount of  $\text{TiO}_2\text{-Pt}$  nanoparticles, respectively much (Fig. 22), or little (Fig. 23). There were not evidenced any ultrastructural alterations of the cell organelles, induced by  $\text{TiO}_2\text{-Pt}$ . Also, in some Kupffer cells, there are many electrondense corpuscles, which probably contain  $\text{TiO}_2\text{-Pt}$  aggregates (Fig. 24).

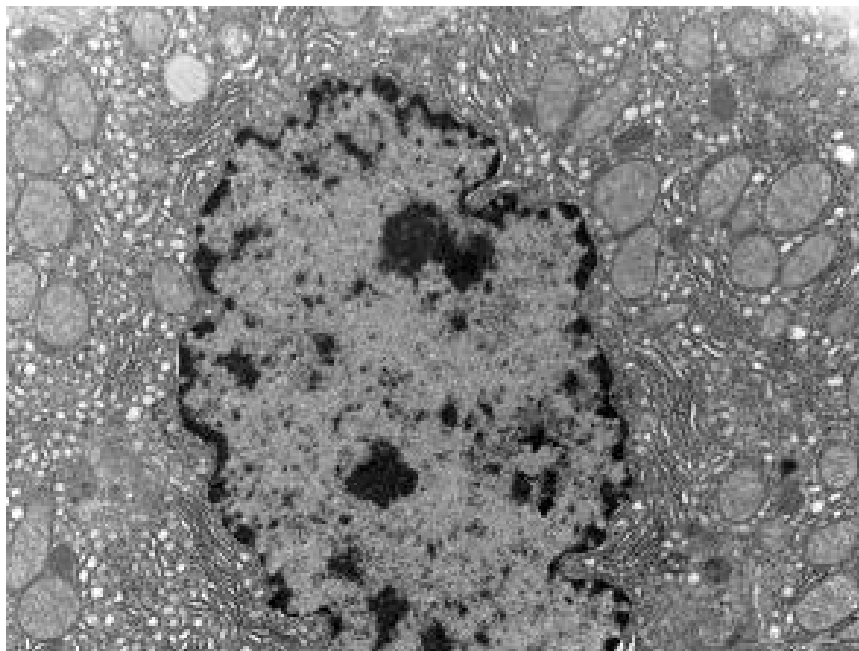


Fig. 20.  $\text{TiO}_2\text{-Pt}$ . Nucleus with unregulated outline.

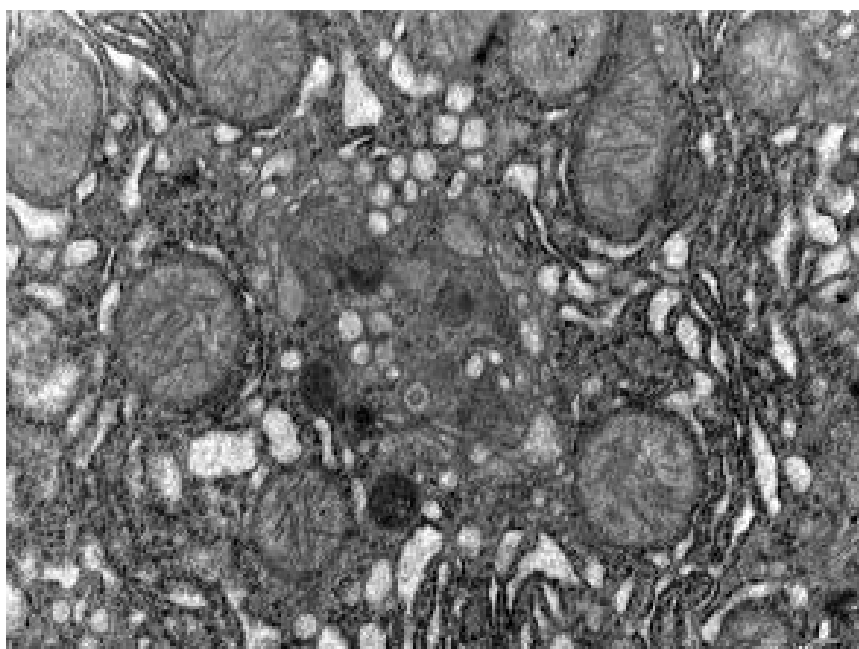


Fig. 21.  $\text{TiO}_2\text{-Pt}$ . Cellular organelles with slightly modifications.

In the hepatocyte of mouse, the  $\text{TiO}_2\text{-Pt}$  nanoparticles, manifest a high reactivity, in comparison with  $\text{TiO}_2\text{-Ag}$  nanoparticles. In the first case, the  $\text{TiO}_2\text{-Pt}$  nanoparticles penetrate in mitochondria and endoplasmic reticulum, while the  $\text{TiO}_2\text{-Ag}$  nanoparticles are present in a

moderate amount in the lipid droplets and massive accumulated in some mitochondria. The analysis of the biological effects of the  $\text{TiO}_2$  doped with different metals (silver, golden, copper or platinum), reveals that the  $\text{TiO}_2$ -Pt and  $\text{TiO}_2$ -Au chelate nanoparticles, manifest an enhanced effects in comparison with  $\text{TiO}_2$ -Ag or  $\text{TiO}_2$ -Cu chelate nanoparticles [9]. Also, the chelated nanoparticles with some metals induced better biological effects in comparison with the unchelated  $\text{TiO}_2$  [9].

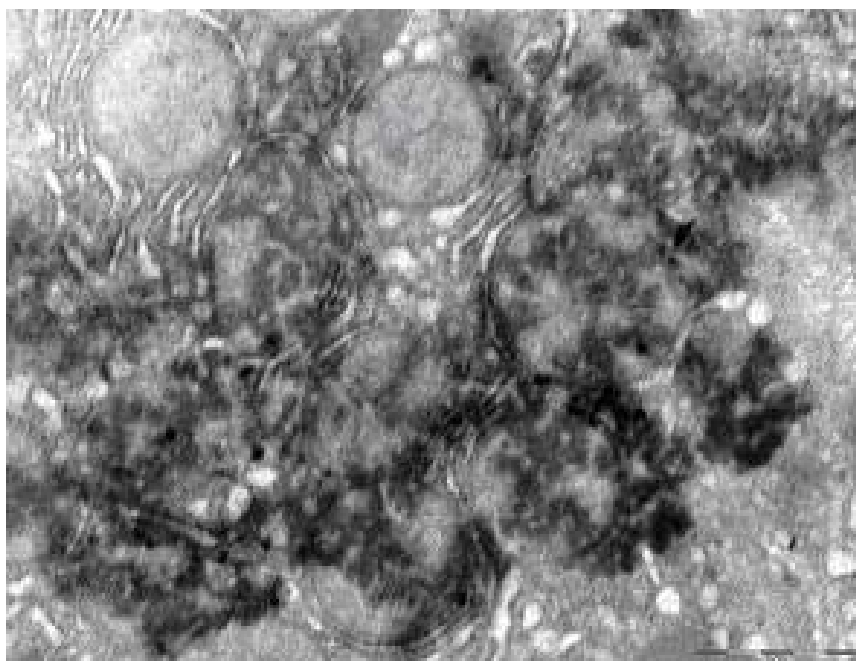


Fig. 22.  $\text{TiO}_2$ -Pt. Nanoparticles in strong relation with mitochondria and RER.

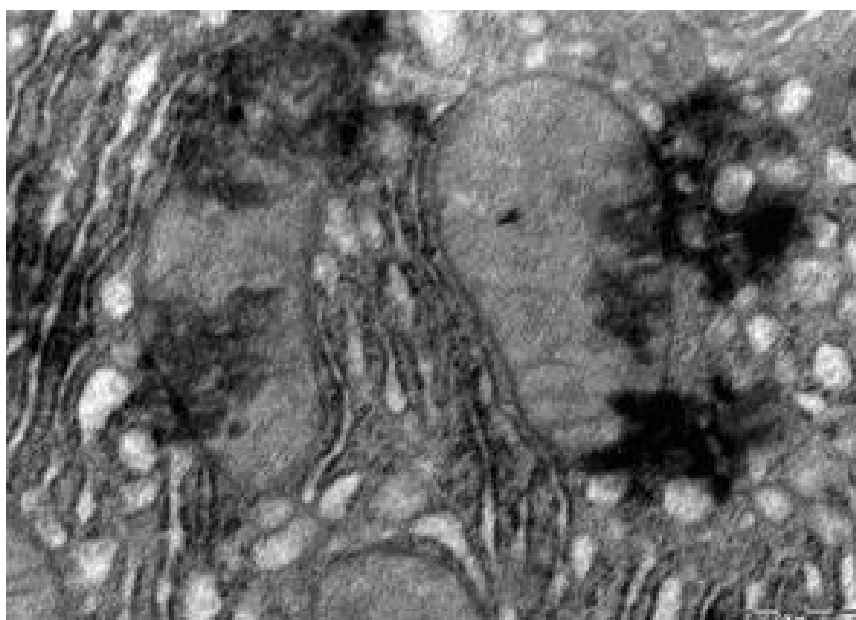


Fig. 23.  $\text{TiO}_2$ -Pt. Nanoparticles in close relations with mitochondria.



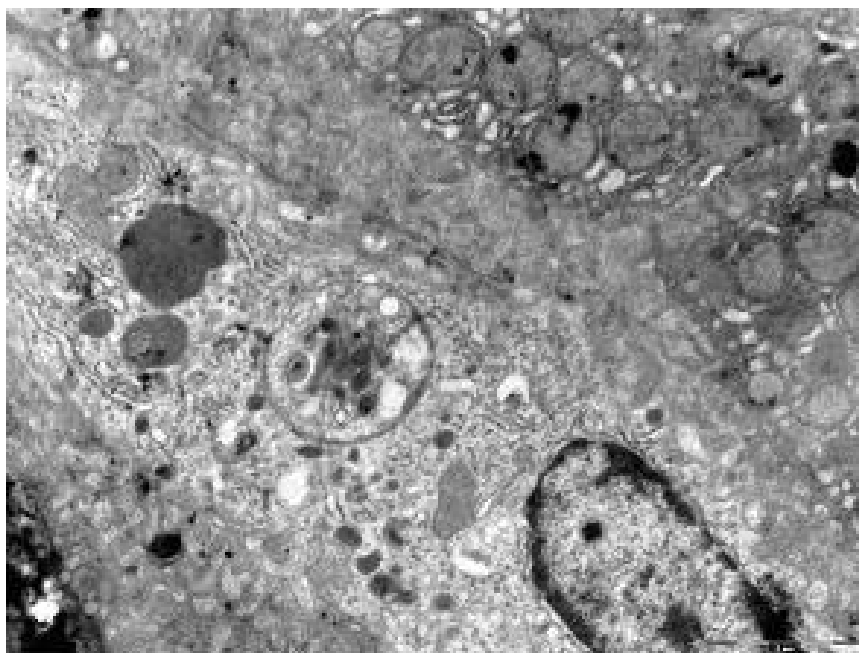


Fig. 24. TiO<sub>2</sub>-Pt. Kupffer cell with electrondense granules.

In some researches performed with TiO<sub>2</sub>-Pt and TiO<sub>2</sub>-Au nanocomposites, point out that these manifest also an enhanced of the photocatalytic reactivity [14]. Also, in other experiment was established that, the chelated nanoparticles with some metals, induced better biological effects in comparison with the unchelated TiO<sub>2</sub> [9]. Similarly, at comparison of the biological effects of the TiO<sub>2</sub> doped with different metals (silver, golden, copper or platinum), the TiO<sub>2</sub>-Pt and TiO<sub>2</sub>-Au chelate nanoparticles, manifest enhanced effects in comparison with TiO<sub>2</sub>-Ag or TiO<sub>2</sub>-Cu chelate nanoparticles [9].

#### **TiO<sub>2</sub>-Fe nanoparticles effect.**

The TiO<sub>2</sub>-Fe nanoparticles induce complex modifications at the hepatocyte level. The nuclei present deep incisures, having an unregulated outline (Fig. 25), the rough endoplasmic reticulum is poor represented, but instead the vesicles of smooth endoplasmic reticulum present a proliferation, as response to this stress factor (Fig. 26). Also, the mitochondria present the matrix and the mitochondrial crista rarefied, with slightly tendency to vacuolization (Fig. 26). The TiO<sub>2</sub>-Fe nanoparticles are present in some hepatocytes, in relation with cellular organelles and nucleus (Figs. 27, 28). The Kupffer cells manifest an increased activity, having in their cytoplasm numerous electrondense granules represented by TiO<sub>2</sub>-Fe and cell remnantes (Fig. 29).

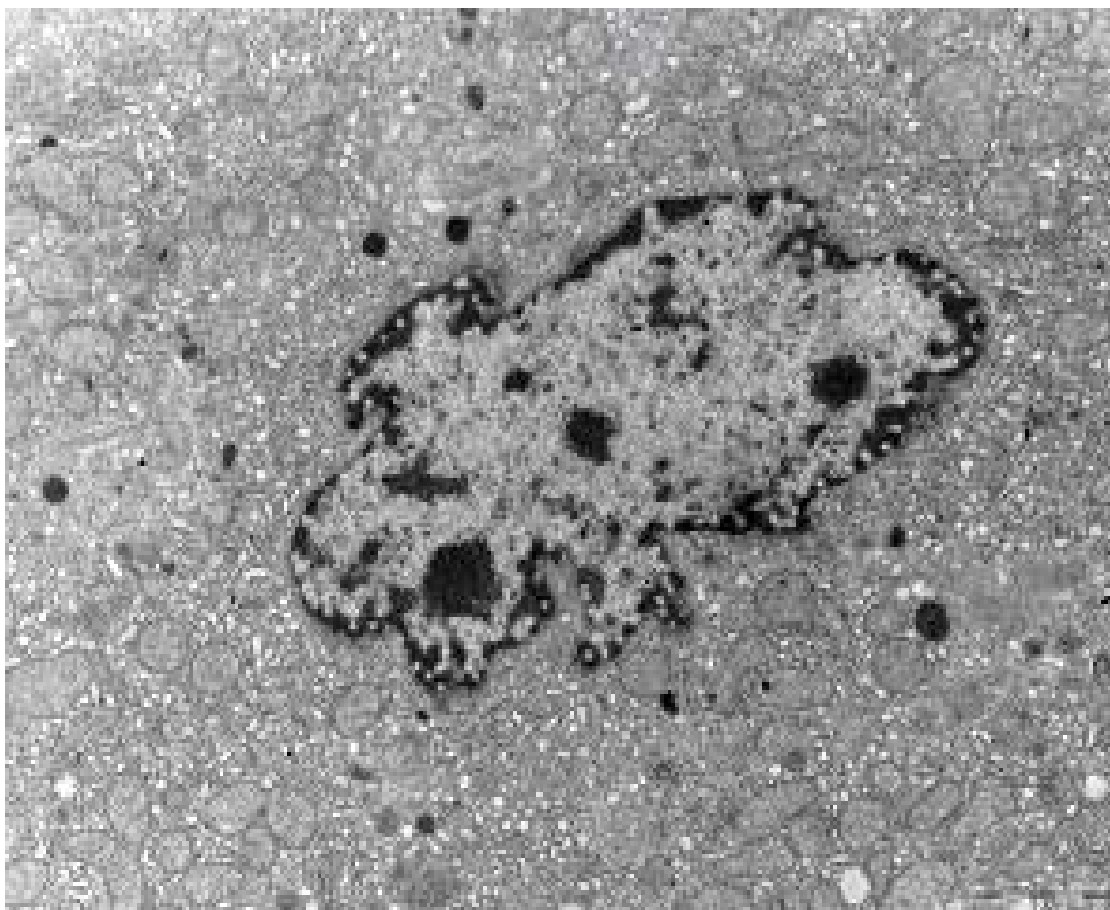


Fig. 25. Nucleus has deep incisures and heterochromatin blocks.

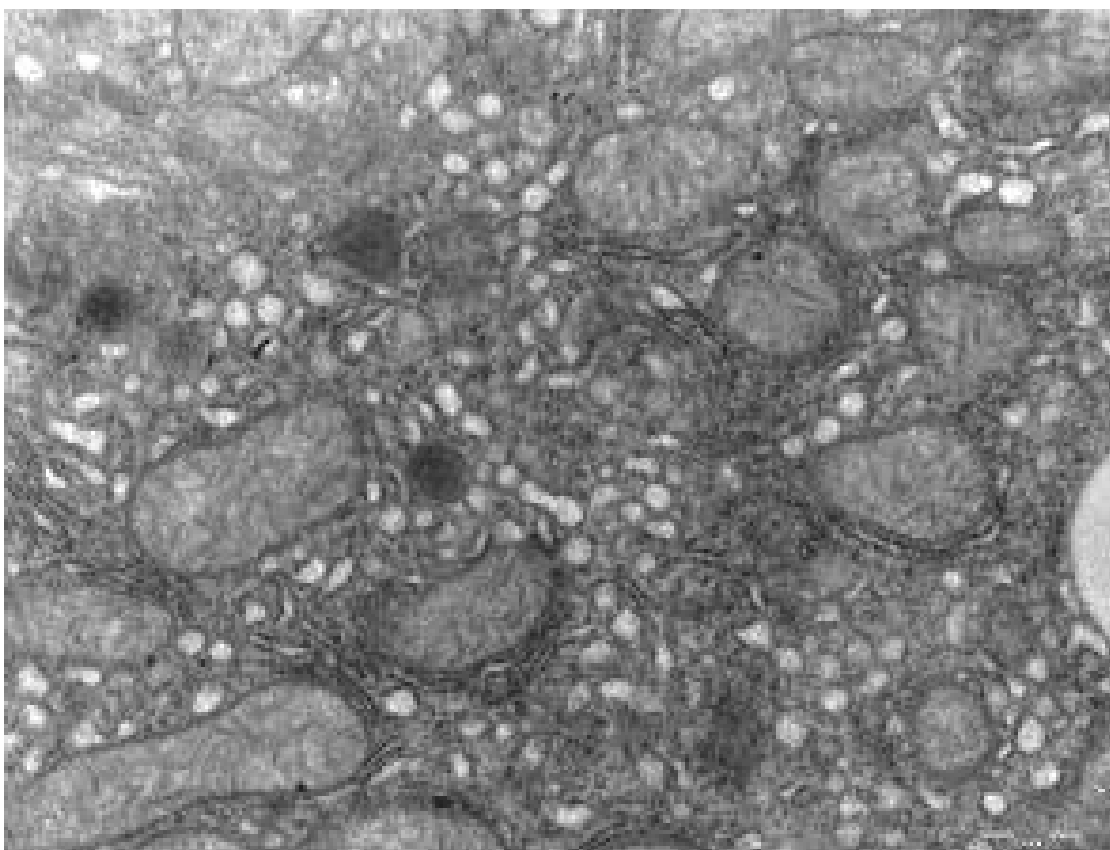


Fig. 26. Cytoplasm with vesicular SER and mitochondria.

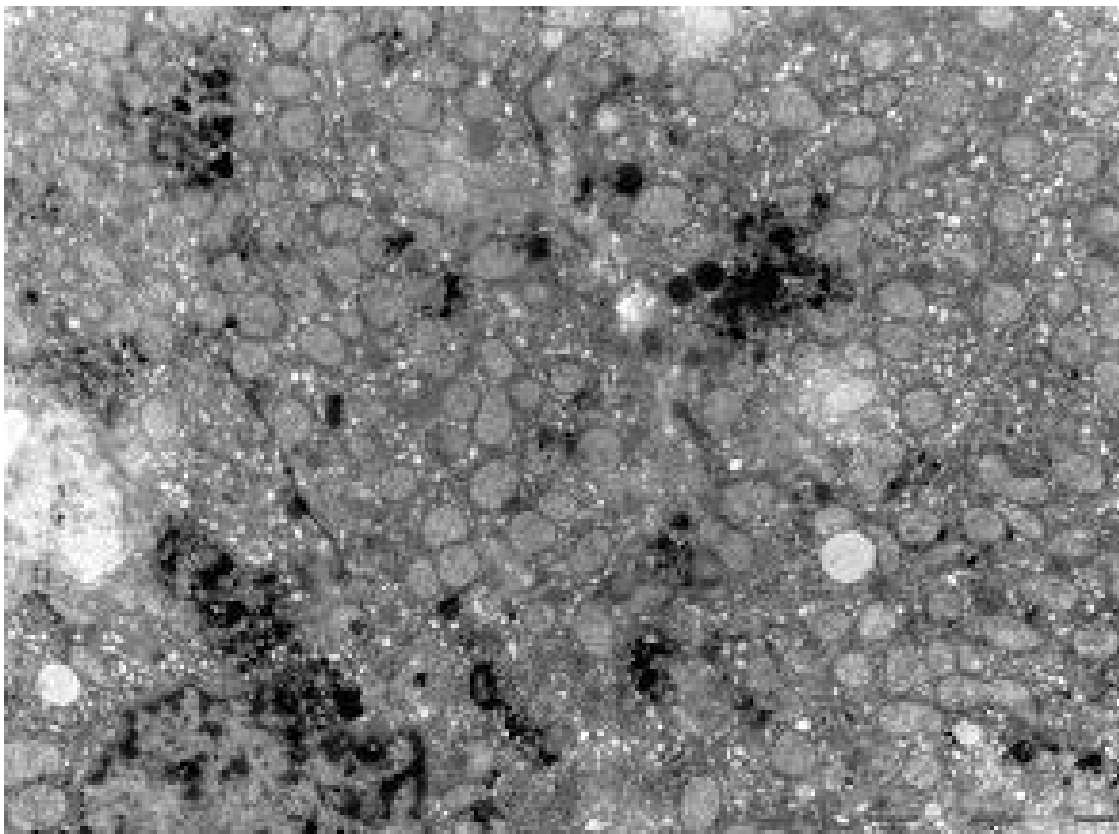


Fig. 27. TiO<sub>2</sub>-Fe nanoparticles in hepatocytes.

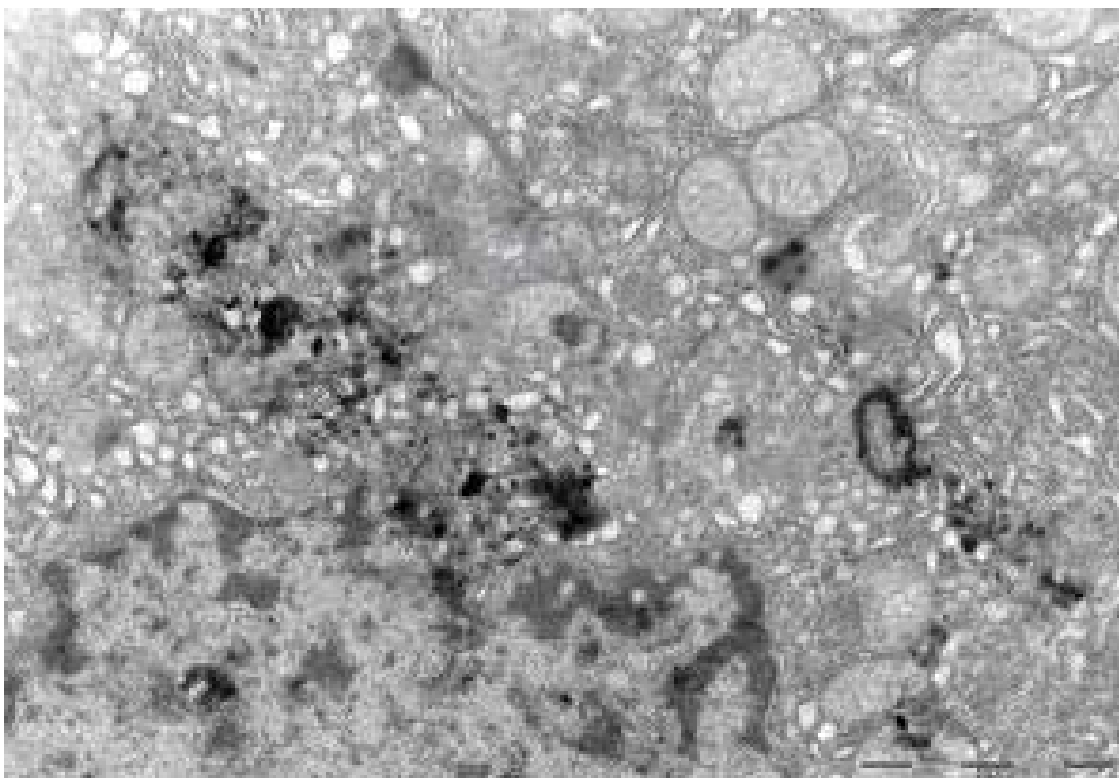


Fig. 28. TiO<sub>2</sub> nanoparticles in relation with nucleus and cellular organelles.

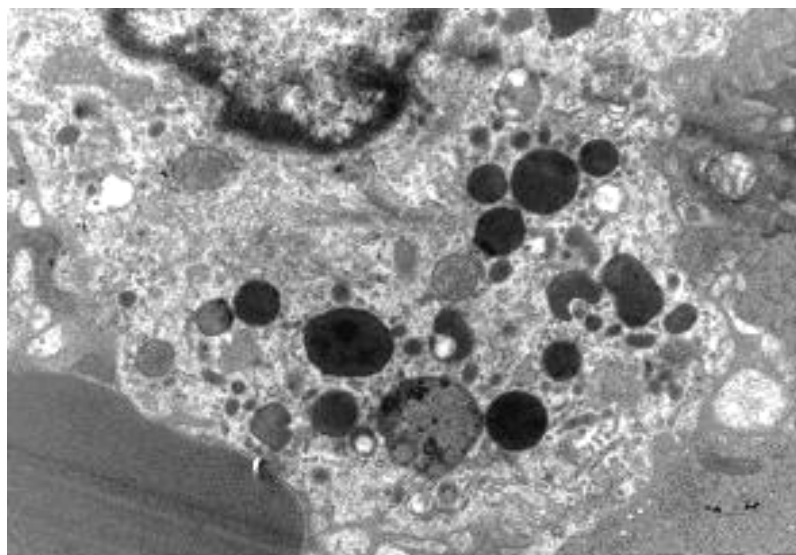


Fig. 29. Kupffer cell with intense activity of phagocytated electrondense bodies

Nanoparticles with iron or iron oxides, present a particular magnetic property (superparamagnetism). In the medical area, they are used for mark and separation of biological materials, deliver of the drugs, analysis with magnetic resonance, a/o.

### Conclusions

In this experiment was analyzed the  $\text{TiO}_2$  nanoparticles effect (*anatase* crystallization form, size of about 10-20 nm), doped or not with a metal, at the liver level in *Mus musculus*, and their interaction with the cellular organelles. The titanium dioxide effect was dependent on the chelated metal. Also, depending on the chelated metal, the  $\text{TiO}_2$  nanoparticles penetrate or not in hepatocyte, and there are in different relations with cellular organelles or cellular inclusions (lipid droplets)

The  $\text{TiO}_2$  nanoparticles, induced a drastic effect at the hepatocyte level, affecting the nucleus and some organelles ultrastructure of organelles (mitochondria especially), as well as the lipid metabolism. The  $\text{TiO}_2$  nanoparticles are accumulated in the lipid droplets, as well as in some mitochondria.

The  $\text{TiO}_2$ -Ag nanoparticles, altered the lipid metabolism, and induced slightly, reversible modifications at the ultrastructural level (dilatation of the rough endoplasmic reticulum, the nucleus outline). In hepatocyte, the  $\text{TiO}_2$ -Ag nanoparticles, are moderate accumulated in lipid droplets, and massive accumulated in mitochondria.

The  $\text{TiO}_2$ -Pt nanoparticles, induced also slightly, reversible modifications. They penetrate in some hepatocytes, being in strong relation with rough and smooth endoplasmic reticulum, and with mitochondria. Their excess is accumulated and degraded in the Kupffer cell.

The  $\text{TiO}_2$ -Fe nanoparticles induced severe alteration of the hepatocytes ultrastructure. They penetrated in some hepatocytes being localized at the rough endoplasmic reticulum and mitochondria level. Their excess was accumulated in the Kupffer cell, as some electrondense bodies.

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